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Article

Introgression of QTLs for Drought Tolerance into Farmers' Preferred Sorghum Varieties

Andekelile Mwamahonje ^{1,2,*}, John Saviour Yaw Eleblu ¹, Kwadwo Ofori ¹, Tileye Feyissa ³, Santosh Deshpande ⁴, Ana Luísa Garcia-Oliveira ⁵, Rajaguru Bohar ⁶, Milcah Kigoni ⁷ and Pangirayi Tongoona ¹

- ¹ West Africa Centre for Crop Improvement (WACCI), College of Basic and Applied Sciences, University of Ghana, Accra 00233, Ghana; jeleblu@wacci.ug.edu.gh (J.S.Y.E.); kofori@wacci.ug.edu.gh (K.O.); ptongoona@wacci.ug.edu.gh (P.T.)
- ² Tanzania Agricultural Research Institute (TARI), Makutupora Centre, P.O. Box 1676, Dodoma 41026, Tanzania
- ³ Institute of Biotechnology, Addis Ababa University, Addis Ababa P.O. Box 1176, Ethiopia; tileye.feyissa@aau.edu.et
- ⁴ International Crops Research Institute for the Semi-Arid Tropics, Patancheru 502324, India; s.deshpande@cgiar.org
- ⁵ Excellence in Breeding (EiB) Platform, International Maize and Wheat Improvement Centre (CIMMYT), ICRAF, Nairobi 00621, Kenya; A.Oliveira@cgiar.org
- ⁶ CGIAR Excellence in Breeding Platform (EiB), CIMMYT, ICRISAT Campus, Hyderabad 502324, India; R.Bohar@cgiar.org
- ⁷ Department of Crop Science, University of Illinois at Urbana-Champaign, Urbana, IL 61801, USA; mkigoni2@illinois.edu
- * Correspondence: andekelilem@gmail.com



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Abstract: Sorghum is a major staple food crop for the people in semi-arid areas of Africa and Asia. Post-flowering drought is a global constraint of sorghum production. The study aimed to improve stay-green (STG) characteristics of farmer-preferred sorghum varieties in Tanzania using marker-assisted backcrossing. A total of 752 individuals representing five BC₂F₁ populations and their parents were genotyped using previously reported KASP markers linked with STG 3A and STG 3B quantitative trait loci (QTL). In the BC₂F₁ populations, the maximum number of individuals with heterozygous alleles were observed in S35*Pato background (37) whereas only seven individuals derived from the B35*Wahi parents' background contained heterozygous alleles. Of the 30 single nucleotide polymorphism (SNP) markers, favourable alleles were observed at 18 loci in BC₂F₁ populations. In the BC₂F₁ generation, the highest (0.127 kg/panicle) grain yield was observed in the B35*NACO Mtama 1 background population. The genotypic analysis revealed the presence of favourable alleles in homozygous conditions at markers loci associated with STG 3A and STG 3B QTLs in BC₂F₃ populations, suggesting successful introgression of STG QTLs from the donor parents to the recurrent parents. Across water irrigation regimes, the highest (0.068 kg/panicle) mean grain weight was observed in the genotype NA316C. Therefore, our study demonstrated the utility of marker-assisted backcrossing for drought tolerance improvement of locally adapted sorghum varieties in Africa.

Keywords: STG; post-flowering drought tolerance; genotyping; single nucleotide polymorphism; marker-assisted selection

1. Introduction

Undoubtedly, drought is a major environmental threat to agriculture sustainability in the world which affects physiological plant growth and performance [1,2]. Nonetheless, both pre-and post-flowering drought stress limits crop yield, though post-flowering drought stress is a major concern for sorghum farmers in Tanzania where it occurs when plants are at the grain filling stage and critical for the determination of yield performance [3,4]. Drought affects water and nutrients uptake from soil and their distribution to

plant parts leading to leaf rolling, leaf senescence and lower vegetative plant growth [4]. Consequently, it affects plant stand, grain size, grain quality and quantity. It is indicated that retention of greenness (referred as 'non-senescence') in leaves during limited water supply accelerates carbon fixation for the starch production [5]. The greenness of leaves allows the photosynthesis process to produce food for the plant and minimises the premature death of flowers, ultimately maintaining yield under drought stress conditions [6]. Generally, farmers escape long dry spells by delaying the planting of crop at the later stage where post-flowering does not fall within the dry spell period [7]; however, recent climate change events make it difficult for farmers and breeders to predict the best time to do it. The common traits indicative of drought tolerance include leaf rolling, leaf senescence, chlorophyll content, days to flowering as well as maturity and root biomass [8]. Moreover, traits such as panicle length, panicle weight, number of seeds per panicle, number of tillers, grain size, dry matter and inflorescence exertion indicate genetic prepotency for drought tolerance [9]. In the past, drought stress in sorghum has been extensively addressed using conventional breeding approaches, but application of modern breeding tools such as genome-wide association, marker-assisted selection (MAS), as well as marker-assisted backcrossing (MABC), is still limited [10]. Nevertheless, drought stress is a highly complex trait, where environmental and crop management practices significantly contribute to the observed variability [10,11] and individual performance. Therefore, the identification of plants that associate correctly with the traits of interest is quite challenging. The exploitation of drought tolerance traits in sorghum had been achieved by mapping STG quantitative trait loci (QTLs) associated with the expression of phenotypic traits such as delay of leaf senescence and maintaining of green leaves at maturity [12]. The efficiency of green leaf area retention depends on the genetic makeup of sorghum genotypes under limited water conditions [5] and governed by traits such as transpiration efficiency, water use efficiency (including water extraction and water use pattern) and leaf canopy development [13]. It is difficult to select plants by phenotyping only because variations may not be due to genetic differences but may be due to environmental effects that may mislead selections [14]. Consequently, researchers have dissected the traits encoding STG [13] and developed molecular markers that are tightly linked to STG QTLs associated with drought tolerance. Based on studies conducted to map STG QTLs in sorghum genotypes SC56, B35, KS19 and E36-1 using different molecular markers, target trait-linked markers were identified [15]. For instance, genotypes B35 and EF36-1 contain novel STG loci such as STG1 and STG2 located on chromosome 3 (SBI-03), STG3 on SBI-02 and, STG4, on SBI-05 which contribute to the expression of the STG phenotypes in sorghum [16]. The genotype B35 sorghum line is characterised by high chlorophyll content, STG and stable root system traits that enable plants to grow and survive better under limited moisture conditions [17]. Moreover, genotypes with STG have high efficiency in utilising nitrogen nutrients from the soil resulting in lower leaf senescence under limited moisture conditions [18,19]. Among the diverse DNA-based markers, SNPs markers clearly detect the existing polymorphism among genotypes, including in sorghum [20,21]. Compared with electrophoresis system-based traditional SNP markers, Kompetitive allele-specific PCRs (KASP) are useful because of low genotyping error rate and amenability to automation that may complement direct phenotyping under field conditions resulting in short breeding cycles in complement with conventional breeding method [2,22]. Additionally, codominant nature of KASP markers facilitates identification of favourable alleles in heterozygous conditions for the introgression of traits through MABC which are either governed by dominant or recessive gene action [23–25]. Previously, two QTLs (STG 3A and STG 3B) expressing leaf greenness under post-drought stress in sorghum have been mapped in 56 to 72 Mbp region of chromosome (SBI-02) for efficiency transpiration and vapour pressure deficit response [26,27]. Therefore, we aimed to introgress both STG 3A and STG 3B QTLs previously described [17,26] from donor parents B35 and S35 to the most important Tanzanian sorghum varieties NACO Mtama 1, Seguifa, Tegemeo, Macia, Pato, Wagita, Wahi, Hakika and Kenya through MABC for diversifying drought tolerance of sorghum varieties.

2. Materials and Methods

2.1. Experimental Materials and Location

The seeds of nine Tanzanian farmers' preferred sorghum varieties, namely Pato, Macia, Seguifa, NACO Mtama 1, Tegemeo, Wagita, Wahi, Hakika and Kenya, were obtained from the Tanzania Agricultural Research Institute (TARI)—Ilonga and Hombolo Centres. The Pato variety originated in Ethiopia but was released in Tanzania after multi-location screening. Pato is a white grain variety that is used for food, and it is characterised by fast recovery from drought, having a maturity period between 95–100 days, highly palatable with hard grain endosperm and no testa. Macia and Seguifa varieties are also white grain and used for food while NACO Mtama1 is white grain used for food and brewing. Tegemeo has white endosperm, cooking quality and cream white grain colour; the grain is used for food and brewing. Wahi and Hakika were released as varieties for tolerance to drought and the *Striga species* in 2002. Wagita and Kenya are local varieties with red-coloured grains, used for food and local alcohol. Wagita and Kenya are late maturing varieties commonly grown in lake zone of Tanzania. Seeds of donor lines B35 and EF36-1 for STG were provided by ICRISAT.

The present study was conducted at TARI-Makutupora Centre located 23 km North of Dodoma city in the Dodoma region of Tanzania (Longitude: 35°, 46.093' E and Latitude: 05°, 58.669' S; Altitude: 1070 m) where rainfalls occur between December and April with a dry spell in February. The site is classified as semi-arid and is characterised by a monomodal rainfall pattern. The annual rainfall at TARI-Makutupora ranges from 300–500 mm with poor distribution and over 500 mm in few years, and the temperature varies from 15–35.1 °C. The crossing and backcrossing were performed in the breeding nursery at TARI-Makutupora from 2017 to 2019 using drip irrigation systems. The field condition for evaluation of BC₂F₃ population performance involved two blocks, the first with well-watered and the second with water stressed at post-flowering. Water stress was imposed from the post-flowering of plants to physiological maturity where no water was applied to plants. Drip irrigation was allowed once a day in well water environment until plant maturity.

2.2. Markers Used for Drought Tolerance

A combined approach of using morphological and molecular markers that track drought tolerance and related traits of agronomic importance was used in this study. This integrated approach made the selection of genotypes of interest effective. The morphological markers used in this study were leaf rolling, leaf senescence, plant height, days to 50% flowering, days to 50% plant maturity, the total number of green leaves at plant maturity, grain colour and grain size, grain weight, root biomass, stem biomass and chlorophyll content were utilised in each generation during field evaluation while molecular markers were integrated at later generations (BC₂F₁ and BC₂F₃ generations) for tracing the favourable alleles at STG 3A as well as STG 3B QTLs, together with some additional markers associated with drought-contributing traits [28,29]. Initially, a total of 30 SNP Kompetitive allele-specific PCR (KASP) markers were used to screen materials [30]. Eight out of these markers were associated with STG 3A and STG 3B, while the remaining twenty-two SNP markers were linked with traits contributing to STG (Table 1). SNP Ids snpSB0035, snpSB0037, snpSB0038, snpSB0072 and snpSB0091 had favourable allele for alternate allele, snpSB0075 and snpSB0080 for recurrent parent allele and the rest had favourable allele for the donor parent allele.

Table 1. List of SNPs markers associated with STG QTLs used for genotyping sorghum.

S.No.	Physi-cal_Mbp	SNP Ids	Alleles	Donor Allele	Alternate Allele	Remarks/Possible
1	S2_56112177	snpSB0035	C/T	C	T	grain types
2	S2_56918230	snpSB0037	C/T	C	T	
3	S2_56918233	snpSB0038	A/T	A	T	
4	S2_57495274	snpSB0039	A/G	A	G	PCD triggering
5	S2_59000770	snpSB0040	C/T	C	T	
6	S2_59047449	snpSB0041	C/G	C	G	
7	S2_59255001	snpSB0042	T/C	T	C	
8	S2_59281892	snpSB0043	T/C	T	C	
9	S2_59472716	snpSB0044	A/T	A	T	
10	S2_59821923	snpSB0049	G/A	G	A	N2 mobilisation/ SAG/SGR1
11	S2_60059010	snpSB0053	A/G	A	G	
12	S2_60098184	snpSB0054	G/A	G	A	
13	S2_61811307	snpSB0072	G/A	G	A	APETALA2 and EREBPs
14	S2_62145285	snpSB0075	A/G	A	G	Recurrent parent-specific
15	S2_62155735	snpSB0076	T/A	T	A	Heat shock domain
16	S2_62155778	snpSB0077	T/C	T	C	
17	S2_62378269	snpSB0080	C/T	C	T	Recurrent parent-specific
18	S2_63375987	snpSB0083	T/C	T	C	Chloroplast precur- sor/ubiquitinosis
19	S2_63690795	snpSB0087	A/G	A	G	
20	S2_65453155	snpSB0089	C/A	C	A	SGR2
21	S2_67306935	snpSB0091	A/C	A	C	panicle compactness
22	S2_67357372	snpSB0092	C/G	G	C	Salt responsive
23	S2_67423749	snpSB0093	A/T	A	T	
24	S2_67664290	snpSB0094	T/C	T	C	
25	S2_67710384	snpSB0095	A/G	A	G	
26	S2_69739036	snpSB0098	C/G	G	C	
27	S2_69859850	snpSB0099	C/T	C	T	
28	S2_70523721	snpSB0101	C/G	C	G	SGR3
29	S2_71360153	snpSB0102	A/G	A	G	
30	S2_71419274	snpSB0103	C/G	G	C	

Of the KASP SNP markers used for genotyping, about 73% and 53% were observed to be polymorphic for the donor and recurrent parents, respectively.

2.3. Introgression of STG QTLs from Donor Parents to the Recurrent Parents and Genotyping

The MABC method was employed to introgress STG QTLs from the donor parents B35 and S35 into the farmers' preferred sorghum varieties (NACO-Mtama 1, Macia, Seguifa, Hakika, Wahi, Pato, Tegemeo, Kenya and Wagita) in Tanzania. The donor parent B35 is drought tolerant and contains STG 1, STG 2, STG 3 and STG 4 QTLs. Parental line S35 is an introgression line derived from B35 as STG QTL donor (STG A). In order to strengthen the stability of drought tolerance in farmers' preferred varieties (recurrent parents), the donor STG alleles were introgressed into recurrent parents by developing backcrossed generations of F_1 , BC_1F_1 , BC_2F_1 , BC_2F_2 and BC_2F_3 . Besides field evaluation, molecular markers were also applied at BC_2F_1 and BC_2F_3 generations for screening STG 3A and STG 3B QTLs.

2.4. The Development of F_1 , BC_1F_1 and BC_2F_1 Populations

A total of 12 crossing plots with 5 rows each were planted with spacing of 0.75 m by 0.30 m (row to row and plant to plant) in two crossing blocks. For crossing, selected plants in recurrent parents were emasculated using the plastic bag method [31]. Pollen was collected in paper bags from the donor parent plants and dusted on the stigma of panicles of the recurrent parents to obtain the F_1 seeds. Pollination bags were removed at the soft dough stage (10–15 days after pollination) and the seed sets on bagged heads were assessed visually using a scale of 0 to 100%, where 0% represented a complete sterile head

without seed set and 100% represented complete seed set. Bird scaring was conducted using tailored small pieces of bags from white mosquito nets which were covered per panicle to prevent birds from eating the seed set after removing the selfing bags.

To generate BC₁F₁ populations, each of the F₁ generations together with their recurrent parents were planted in single row plot of 5.4 m long and were backcrossed to their respective recurrent parents. Similarly, BC₁F₁ seeds were used to generate subsequent BC₂F₁ populations.

2.5. Development of BC₂F₂ and BC₂F₃

For generation of BC₂F₂ population, a total of five genotypes comprising of three genotypes with heterozygous alleles (W82, NA241 and NA307) and two genotypes with homozygous alleles (NA316 and SE438) from the BC₂F₁ population were selected following genotyping analysis and agronomic performance under field conditions. Subsequently, genotypes NA241A, NA241B, NA316A, NA316B, NA316C, NA307, SE408 and SE438 were screened from the population BC₂F₂ as the best-performing under field conditions to generate the BC₂F₃ population.

2.6. Genotyping of BC₂F₁ and BC₂F₃ Populations

For introgression of STG QTLs, genotyping of BC₂F₁ populations was performed with a KASP marker (LGC, Middlesex, UK) in an agreement with Intertek-AgriTech (<https://www.intertek.com/agriculture/agritech/>: accessed on 20 July 2021) as an external service provider [32]. A total of 752 samples representing 150 individual leaf samples per BC₂F₁ population, together with donor and recurrent parents, were collected in the 96-well plates. Three-leaf discs of approximately 4–5 cm per plant sample were collected using single-hole punching methodology. Following genotyping of the BC₂F₁ population, the presence of favourable alleles at STG loci in the BC₂F₃ population was also confirmed with ten KASP markers tightly linked to STG 3A and STG 3B QTLs (SnpSB0042, SnpSB0049, SnpSB0053, SnpSB0054, SnpSB0072, SnpSB0089, SnpSB0098, SnpSB0101, SnpSB0102 and SnpSB0103). Seven BC₂F₃ populations (NA241A, NA241B, NA316A, NA316B, NA316C, SE408 and SE438) were chosen and at least 100 samples per populations together with parents, making a total of 752 samples, were genotyped. The genotype NA307 was evaluated under field conditions only following the limitation of samples needed for genotyping. The SNP markers used for genotyping at BC₂F₁ and BC₂F₃ populations are indicated in Table 1.

2.7. Molecular Data Analysis

Marker data analysis was performed using SNPviewer from Biosearch Technologies (<https://www.biosearchtech.com/support/tools/genotyping-software/snpviewer>, LGC, Middlesex, UK: accessed on 25 December 2020) for insurance of marker quality. Subsequently, genotyped lines were compared in flajack v. 1.21.02.04 (<https://ics.hutton.ac.uk/flajack/>: accessed on 25 December 2020).

3. Results

3.1. Phenotypic Data

Five percent of the best-performing genotypes of the BC₂F₁ populations for the traits plant height, STG, total number of green leaves at maturity, panicle length, panicle width and panicle weight that were screened under field conditions showed significance differences of performance in all traits evaluated across replication. The highest mean plant height (146.4 cm) was recorded in the population S35*Pato (Table 2), whereas donor parent B35 exhibited the lowest (88 cm) plant height. The selected 5 percent BC₂F₁ population performed better than their parents. The population B35*NACO Mtama 1 showed the highest grain yield (0.127 kg/panicle), while the donor parent B35 produced the lowest (0.013 kg/panicle). The best-performing BC₂F₁ genotypes under field conditions and those identified to be desirable at a genotypic level were screened for further improvements.

Table 2. Five per cent of the best performance BC₂F₁ populations and their parents.

Population	PH	STG	NGLM	PL	PW	EX	GWT
B35*Wahi	117.4 ^c	4.83 ^{ab}	2.167 ^{ef}	32.22 ^f	7.5 ^{fg}	2.5 ^b	0.096 ^e
S35*Hakika	135 ^{defg}	5.5 ^{cd}	2 ^{de}	30.25 ^e	7.25 ^{ef}	2 ^a	0.08 ^d
B35*NACO	144.1 ^{gh}	4.67 ^a	3 ^g	24.92 ^d	8.17 ^g	3 ^c	0.127 ^f
B35*Macia	132.7 ^{def}	6.67 ^f	1 ^{bc}	23.88 ^d	5.07 ^{cd}	3 ^c	0.027 ^a
B35*Seguifa	141.7 ^{fgh}	5 ^{abc}	2.833 ^{fg}	20.75 ^c	6.58 ^e	2.83 ^{bc}	0.11 ^e
S35*Pato	146.4 ^h	6.17 ^{ef}	1 ^{bc}	21.75 ^c	8.17 ^g	2.5 ^b	0.112 ^e
B35 DP	88 ^a	8 ^g	0.33 ^{ab}	17.17 ^{ab}	3.5 ^a	3 ^c	0.013 ^a
Pato RP	131.3 ^{de}	5 ^{abc}	1.67 ^{cde}	15.83 ^a	6.67 ^e	2.67 ^{bc}	0.064 ^c
Seguifa RP	139.7 ^{efgh}	5.33 ^{bcd}	1.67 ^{cde}	21.33 ^c	5.67 ^d	2.67 ^{bc}	0.067 ^{cd}
Hakika RP	131.2 ^{de}	5.33 ^{bcd}	1 ^{bc}	29.17 ^e	5.5 ^d	3 ^c	0.028 ^a
Macia RP	113.4 ^{bc}	7.67 ^g	0.0 ^a	21.33 ^c	4 ^{ab}	3 ^c	0.019 ^a
NACO RP	113.8 ^{bc}	5.67 ^{de}	1.33 ^{cd}	21.47 ^c	5.13 ^{cd}	3 ^c	0.047 ^b
S35 DP	104.5 ^b	5.67 ^{de}	2 ^{de}	18.33 ^b	4 ^{ab}	3 ^c	0.024 ^a
Wahi RP	128.3 ^d	5 ^{abc}	1 ^{bc}	30.17 ^e	4.5 ^{bc}	3 ^c	0.021 ^a
SE	2.72	0.16	0.22	0.5	0.2	0.11	4.43
LSD	5.37	0.32	0.43	0.98	0.4	0.33	8.72
CV (%)	6.5	8.4	43.2	6.3	10.5	11.7	22.3

^{a-h} are compared by Bonferroni mean test; CV (%)—Coefficient of variation of means; DP—Donor parent. EX—Inflorescence exersion; GWT—Grain weight (g); LSD—Least significance difference of means; NACO—NACO Mtama 1; NGLM—Total number of green leaves at maturity; PH—Plant height (cm); PL—Panicle length (cm); PW—Panicle width (cm); RP—Recurrent parent; SE—Standard error of means; STG—Stay-green (Scale: 1—very low or no visible sign of susceptibility, 9—very high).

3.2. BC₂F₁ Genotyping

Of the 728 BC₂F₁ samples collected for genotyping, about 10% samples (71 plants) were observed to be heterozygous as representative SNP markers, detailed in Table 3. Of these, three (snpSB00075, snpSB00102 and snpSB00103) and two (snpSB00102 and snpSB00103) SNPs markers were scored as heterozygous in a total of seven and eight samples with BC₂F₁ of the B35*Wahi and BS35*Hakika backgrounds, respectively. Similarly, heterozygous alleles at three (snpSB00072, snpSB00077 and snpSB00103) and four markers (snpSB00049, snpSB00077, snpSB00102 and snpSB00103) loci were also identified in 37 samples of S35*Pato and 10 samples of BC₂F₁ with the B35*Seguifa background. The markers snpSB00037, snpSB00041, snpSB00098, snpSB00102 and snpSB00103 showed heterozygous alleles in 10 samples with B35*Macia backgrounds, respectively. The markers snpSB00042, snpSB00098, snpSB00102 and snpSB00103 expressed heterozygous alleles in a total of 18 samples of the B35*NACO Mtama 1 background (Table 3). The rest (19) of the SNP markers showed homozygous alleles of the BC₂F₁ samples used for genotyping. The complete details of markers are shown in the Supplementary Table S1 in the main text connecting Table 3 data.

Eighteen SNP markers showed favourable alleles among 728 of BC₂F₁ samples genotyped (Table 4). The markers that indicated favourable alleles that were most important for STG included: snpSB00049 and snpSB00054 for STG 3A, and snpSB00102 and snpSB00103 for STG 3B.

Table 3. BC₂F₁ sorghum populations scored with heterozygous alleles using SNP markers.

NS	SNP Markers	H. Allele	DP Alleles	RP Alleles	NS	SNP Markers	H. Allele	DP Alleles	RP Alleles
BC ₂ F ₁ (B35*Wahi)					BC ₂ F ₁ (B35*Seguifa)				
82	snpSB0102	G/A	AA	GA	567	snpSB0103	G/C	CC	CC
90	snpSB0102	G/A	AA	GA	569	snpSB0103	G/C	CC	CC
94	snpSB0102	G/A	AA	GA	581	snpSB0103	G/C	CC	CC
BC ₂ F ₁ (S35*Hakika)					BC ₂ F ₁ (S35*Pato)				
127	snpSB0103	G/C	CC	CC	595	snpSB0049	G/A	GG	AA
131	snpSB0102	G/A	AA	GA	601	snpSB0103	G/C	CC	CC
138	snpSB0103	G/C	CC	CC	610	snpSB0049	G/A	GG	CC
BC ₂ F ₁ (NACO Mtama 1)					624	snpSB0102	G/A	AA	AA
213	snpSB0103	G/C	CC	CC		snpSB0103	G/C	GG	CC
276	snpSB0103	G/C	CC	CC	645	snpSB0102	G/A	AA	AA
304	snpSB0102	G/A	AA	AA	656	snpSB0103	G/C	CC	CC
307	snpSB0102	G/A	AA	AA	659	snpSB0102	G/A	AA	AA
319	snpSB0103	G/C	CC	CC		snpSB0103	G/C	CC	CC
320	snpSB0103	G/C	CC	CC	663	snpSB0103	G/C	CC	CC
BC ₂ F ₁ (B35* Macia)					668	snpSB0103	C/T,G/C	CC	CC
347	snpSB0103	G/C	CC	CC	671	snpSB0103	G/C	CC	CC
348	snpSB0102	G/A	AA	AA	679	snpSB0103	G/C	CC	CC
352	snpSB0103	G/C	CC	CC	685	snpSB0102	G/A	AA	AA
391	snpSB0103	G/C	CC	CC		snpSB0103	G/C	CC	CC
395	snpSB0103	G/C	CC	CC	703	snpSB0102	G/A	AA	AA

H—Heterozygous; DP—Donor parent; NS—Number of samples; RP—Recurrent parent.

Table 4. Favourable alleles and STG QTLs identified after genotyping BC₂F₁ populations of sorghum.

SNP Markers	FA	QTL	Physical Location	Role
snpSB00039	A	STG 3A	S2_57495274	Retains STG and triggers PCD
snpSB00040	C	STG 3A	S2_59000770	Retains STG and triggers PCD
snpSB00041	C	STG 3A	S2_59047449	Retains STG and triggers PCD
snpSB00043	T	STG 3A	S2_59281892	Retains STG and triggers PCD
snpSB00044	A	STG 3A	S2_59472716	Retains STG and triggers PCD
snpSB00049	G	STG 3A	S2_59821923	Retains STG and enhances grain yield
snpSB00053	A	STG 3A	S2_60059010	Retains STG and enhances grain yield
snpSB00076	T	STG 3A	S2_62155735	Retains STG and regulate heat shock domain
snpSB00077	T	STG 3A	S2_62155778	Retains STG and regulate heat shock domain
snpSB00083	T	Interval	S2_63375987	Retains STG and regulates chloroplast precursors
snpSB00087	A	Interval	S2_63690795	Retains STG and regulates chloroplast precursors
snpSB00089	C	STG 3B	S2_65453155	Retains STG and enhances grain yield
snpSB00093	A	STG 3B	S2_67423749	Retains STG and regulates salt
snpSB00094	T	STG 3B	S2_67664290	Retains STG and regulates salt
snpSB00095	A	STG 3B	S2_67710384	Retains STG and regulates salt
snpSB000101	C	STG 3B	S2_70523721	Retains STG and enhances grain yield
snpSB00102	A	STG 3B	S2_71360153	Retains STG and enhances grain yield
snpSB00103	G	STG 3B	S2_71419274	Retains STG and enhances grain yield

SNP—Single nucleotide polymorphism; FA—Favourable alleles; QTL—Quantitative trait loci; STG—Stay-green; PCD—programmed cell death.

In order to generate BC₂F₃ populations, three samples (W82, NA241 and NA307) showing heterozygous alleles and two samples (NA316 and SE438) with homozygous favourable alleles for STG QTL were chosen. The favourable alleles at markers associ-

ated with STG 3B QTL were found in plant W82 (snpSB00102), NA241 (snpSB00102 and snpSB00103) and NA307 (snpSB00101 and snpSB00102). Moreover, plant sample NA316 had favourable alleles in homozygous conditions for SNP markers associated with STG 3A (snpSB00049, snpSB00053 and snpSB00054) and STG 3B (snpSB00102). Similarly, plant SE438 also had homozygous alleles of SNP marker snpSB00054 and snpSB00102 associated with STG 3A and STG 3B, respectively. Plants which expressed heterozygous alleles after genotyping were categorised as the priority because of the availability of traits from both parents for successful STG introgression from either of the donor parents B35 or S35 to the recurrent parents. Prior to selection, phenotypic data were recorded for traits such as days to 50% flowering, plant height, days to 50% plant maturity, panicle weight, grain weight per plant, seed vigour, stability of plants, leaf senescence, susceptible to pests and diseases. Plants with good performance based on phenotypic data, such as days to 50% flowering, plant height, days to 50% plant maturity, panicle weight, grain weight per plant, seed vigour, stability of plants, leaf senescence and susceptibility to pest and diseases, were selected and compared with genotyped data. Finally, best-performing plants under field conditions with favourable alleles for STG 3A and STG 3B were selected.

3.3. Phenotypic Data of BC₂F₃ Populations' Results

The mean grain weight per panicle evaluated across water irrigation and water stress condition was the highest (0.068 kg) in the genotype NA316C followed by NA307 with 0.064 kg (Figure 1). The lowest (0.039 kg) mean grain yield was recorded in the donor parent B35. The mean grain weight of the rest of the genotypes ranged from 0.049–0.061 kg.

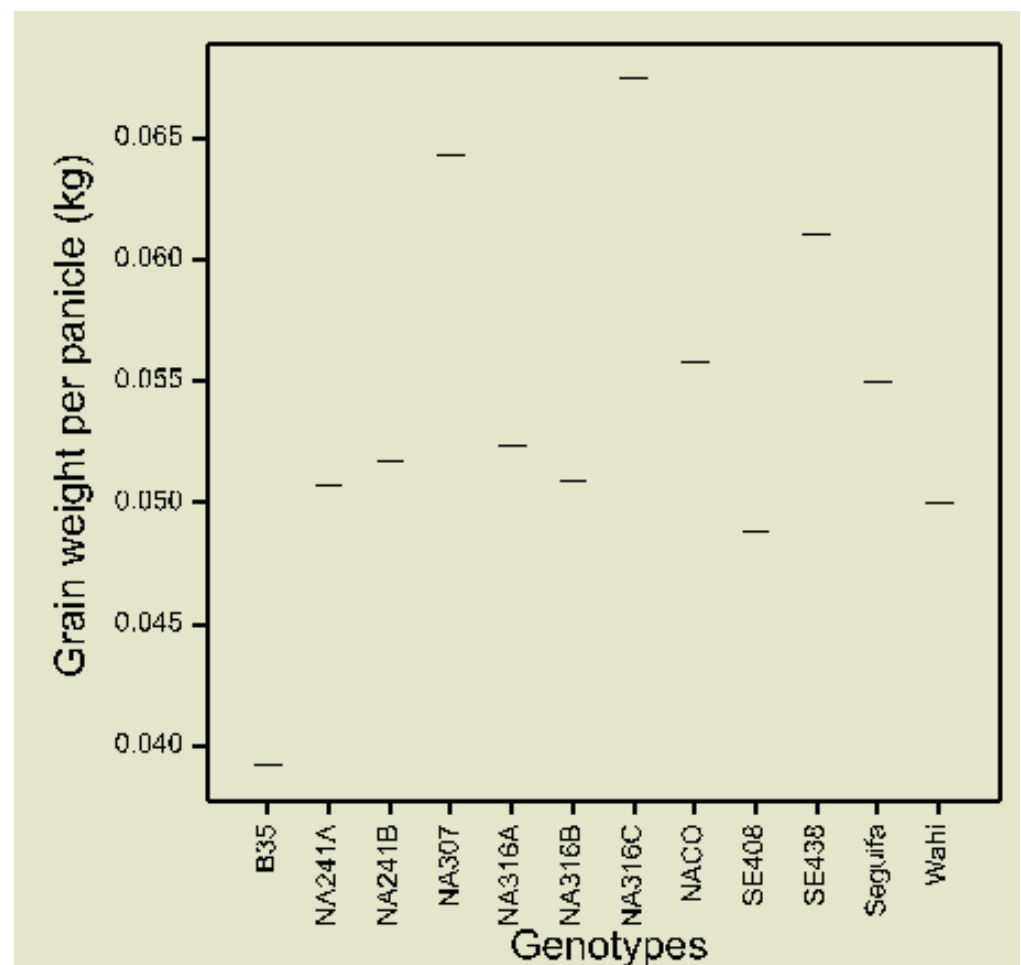


Figure 1. Boxplot of mean grain weight per panicle of BC₂F₃ genotypes and parents across well-watered and water-stressed environments.

The mean STG per genotype screened across water regimes was scored the highest (4.45) in the donor parent B35 next by the control Wahi with 4.7 score (Figure 2). The lowest (6.23) score was recorded by the genotype NA307. However, of the BC₂F₃ populations, the genotype NA316A showed the highest STG retention in sorghum leaves. No significance differences were recorded in the rest of the genotypes.

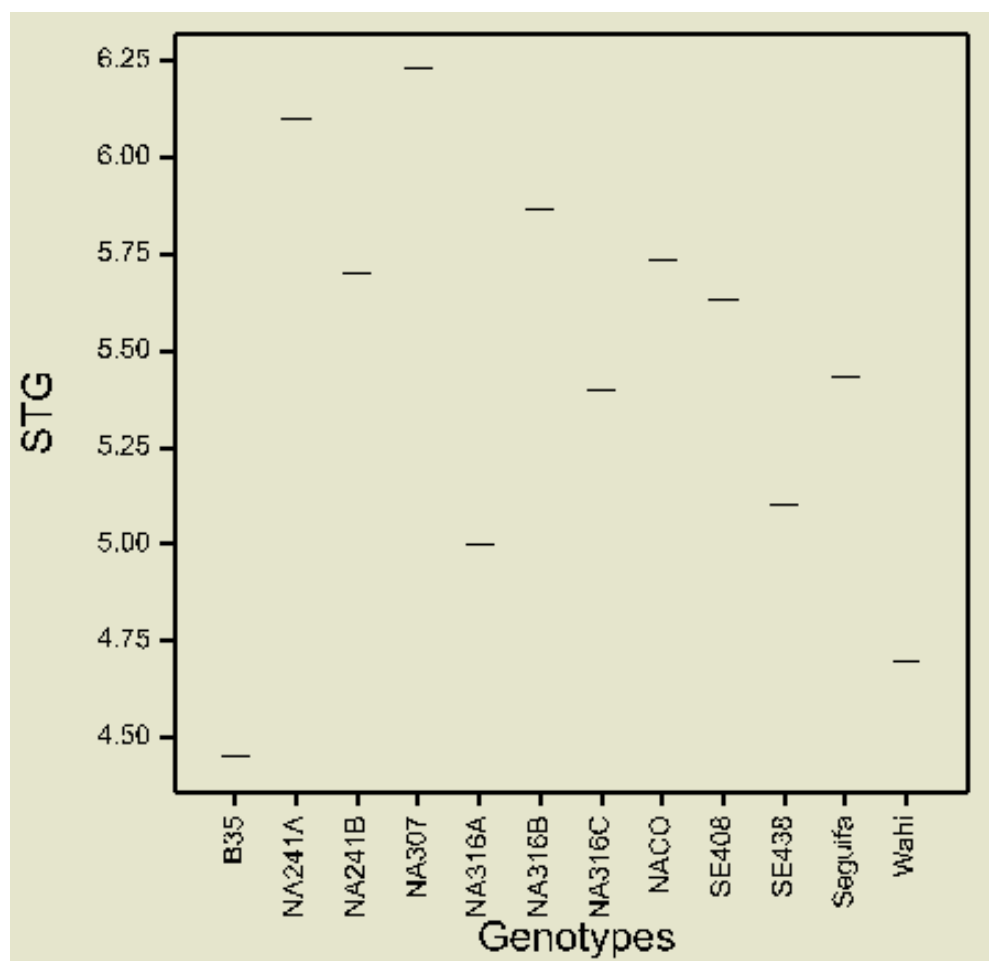


Figure 2. Boxplot of mean STG of BC₂F₃ genotypes and parents across well-watered and water-stressed environments (Scale: 1–9 where, 1—all leaves in plant are green, 9—all leaves in plant are dried).

3.4. BC₂F₃ Genotyping Results

Eight out of ten SNP markers (snpSB0042, snpSB0049, snpSB0053, snpSB0054, snpSB0072, snpSB0089, snpSB0101 and snpSB0102) used for genotyping of BC₂F₃ population identified favourable homozygous alleles for STGs in sorghum as in the representative SNP marker details in Table 5. The SNP markers snpSB0042, snpSB0049, snpSB0053, snpSB0054, snpSB0072 and snpSB0089 identified favourable homozygous alleles for STG 3A QTL. The snpSB0101 and snpSB0102 markers linked favourable alleles for STG 3B QTL (Table 5). The SNP markers, snpSB00042 identified the lowest (2) number of favourable alleles in the donor parent B35. The snpSB00098 and snpSB0103 markers did not link favourable alleles in the population genotyped. The SNP markers, snpSB00054, snpSB00089 and snpSB00101, recorded the highest number (124) of samples with favourable alleles in almost all of the genotypes analysed. SNP markers which failed to score the favourable alleles and STG 3A and STG 3B QTLs were not included in Table 5. The complete details are shown in the Supplementary Table S2 in the main body text connecting Table 5 data.

Table 5. Genotypes of BC₂F₃ populations scored by favourable alleles using SNP markers.

Genotype	SNP Markers	FA	No. of FA	Genotype	SNP Markers	FA	No. of FA
BC ₂ F ₃ (B35*NACO)				BC ₂ F ₃ (B35*Seguifa)			
NA241A	snSB00049	GG	115	SE408	snSB00049	GG	55
	snSB00072	AA	6		snSB00089	CC	73
	snSB00089	CC	124		snSB00101	CC	73
	snSB000102	AA	123	SE438	snSB00049	GG	23
NA241B	snSB00049	GG	113	Seguifa (Recurrent P)	snSB00053	AA	51
	snSB00101	CC	122		snSB000102	AA	52
NA316A	snSB00049	GG	116	B35 (Donor P)	snSB00042	TT	2
	snSB00053	AA	120		snSB00049	GG	2
	snSB00101	CC	121		snSB00101	CC	2
NA316B	snSB00049	GG	70	Wahi	snSB000102	AA	2
	snSB00053	AA	115		snSB00103	GG	2
	snSB000102	AA	115		snSB00049	GG	3
NA316C	snSB00072	AA	27	snSB00101	CC	2	
	snSB000102	AA	121				
	snSB00049	GG	92				
NACO Mtama 1 (Recurrent P)	snSB00053	AA	3				
	snSB00054	GG	3				

Donor P—Donor parent, Recurrent P—Recurrent parent.

The SNP marker snpSB00089 indicated the highest (729) total number of samples with favourable homozygous alleles for STG 3B followed by snpSB00101 (728; Table 6). The SNP markers which failed to show favourable alleles linked to STG QTLs such as snpSB00098 and snpSB00103 were not included in Table 6. The SNP markers snpSB00042, snpSB00049, snpSB00053, snpSB00054, snpSB00072 and snpSB00089 were associated with STG 3A QTL. The snpSB0101 and snpSB0102 markers were linked with STG 3B QTL in the BC₂F₃ populations.

Table 6. Favourable alleles and STG QTLs identified in BC₂F₃ populations of sorghum.

SNP Markers	FA	No. of FA	QTL	Physical Location	Function
snpSB00042	T	2	STG 3A	S2_59255001	Retain STG and PCD triggering
snpSB00049	G	584	STG 3A	S2_59821923	Retain STG and N mobilisation
snpSB00053	A	707	STG 3A	S2_60059010	Retain STG and N mobilisation
snpSB00054	G	709	STG 3A	S2_60098184	Retain STG and N mobilisation
snpSB00072	A	105	STG 3A	S2_61811307	EREBPs
snpSB00089	C	729	STG 3B	S2_65453155	Retain STG
snpSB00101	C	728	STG 3B	S2_70523721	Retain STG
snpSB00102	A	727	STG 3B	S2_71360153	Retain STG

EREBPs—ethylene-responsive element binding proteins; FA—Favourable allele; No. FA—Number of favourable allele; N—Nitrogen; PCD—Programmed cell death; QTL—Quantitative trait loci, SA—salicylic acid; SNP—Single nucleotide polymorphism; STG—stay-green.

4. Discussion

Findings in this study revealed the importance of molecular markers to shorten breeding cycles in sorghum. The genotyping information is crucial for exploiting traits from the parents to improve new lines, as STG is a complex trait associated with several genes. The introgression of STG QTLs by MABC enhances post-flowering drought tolerance in sorghum [4]. The transfer of the STG trait from the donor parents B35 and S35 into the farmers' preferred varieties using SNP markers is highly desirable for post-flowering drought tolerance. This study identified favourable alleles for donor and recurrent parents on STG 3A and STG 3B in the samples of the genotype BC₂F₁ and BC₂F₃ populations, indicating successful introgression. The incorporation of STG QTLs improves plants' delayed leaf senescence from post-flowering to physiological plant maturity [12]. Sukumaran et al. [33] reported 8–24% phenotypic variations contributed by STG QTLs with the flanking SNP markers. Other studies which used SSRs markers for improvement of STG 1, STG 2, STG 3

and STG 4 in sorghum reported successful introgression [34,35]. The SNP markers indicate the arrangement of bialleles in the whole genome. The markers identify alleles with a high rate of recovery in the genome, which make their precision higher than other markers [30]. Findings in the current study show that some of the new genotypes generated by the backcrossing method performed well in grain yield compared with control and recurrent parents planted on the same date under similar environments. The outperformance of new genotypes over farmers' cultivation varieties shows the advantages of SNP markers for high efficiency STG QTLs in sorghum improvement. STG 1 QTL mapped by SSR markers adapts post-flowering drought stress and it increases yield grain more than other QTLs, thus it is named as the best trait of the SSR markers [1,36]. However, QTLs STG 3A and STG 3B introgression to farmers' drought susceptible cultivated sorghum varieties have reported significant improved efficiency of green area retention at post-flowering drought stress [37]. The QTLs have improved grain yield and water use efficiency at physiological plant maturity, making SNP the best markers for mapping tightly linked QTLs for STG compared with SSRs in the post-drought tolerance improvement in sorghum [26,37]. Currently, SNPs markers are the most commonly used markers for identification of genetic polymorphism which are used in the study of genetic variation among populations [38,39]. Da Silva et al. [30] used two backcrosses to introgress the *bmr6* allele for biomass from the donor parent CMSXS170 line to elite lines CMSXS652 and IS23 of sweet sorghum using SNP markers. Results revealed a successful transfer of the *bmr6* allele to the recurrent parent for biomass improvement. For successful backcrosses, the promising genotypes selected for traits of interest should contain favourable alleles from either one of the parents and must be expressed phenotypically [12]. Backcrosses must be done at least three to four times for sufficient transfer of the QTLs from the donor to the recurrent parents, followed by successive selfing until the backcross line is close to the original recurrent parent [30]. SNPs markers narrow the specific location of QTLs which is closely linked to the gene expressing the STG trait. The SNP markers identify favourable alleles for STG expression, thus hastening screening of drought tolerance sorghum genotypes. The current study identified favourable alleles for STG that were screened for further studies of sorghum improvement. It was noted that some of the plants performed well in the field environments by expressing STG, low leaf rolling, high yield and high vigour of sorghum grains, but were not tightly linked by SNP markers used for genotyping. This indicates that there is a need to map a large number of SNP markers to widen the chance of identifying the right STG QTLs. If SNP markers fail to locate the position of the QTL expressing STG in the chromosomes, SSR and DArT markers can alternatively be used [32]. Moreover, findings have reported that there are some QTLs which are detected by molecular markers in the backcrosses but fail to express under field conditions; such genotypes do not qualify for selection of crop improvement [40]. Differential transcriptome analysis encompasses bioprocess to map genes controlling the expression of drought tolerance in sorghum [41]. The STG expression on drought tolerant sorghum varieties is due to the reaction of genes which increase the production of antioxidant capacity, regulatory factors and the repressors of early senescence [41]. The application of modern technology supplements conventional breeding techniques to address drought stress in sorghum. With modern technology, for instance, molecular markers map traits of interest which saves time of breeding cycles. In this study, SNP markers identified favourable alleles for STG QTLs from the genotypes of BC₂F₁ and BC₂F₃ populations which simplified the comparison with field data to screen the best plants with the target traits. It is important to use markers that are powerful and reliable to identify the favourable alleles for STG, QTLs and chromosome position of genotypes for inclusion in drought tolerance sorghum improvement.

5. Conclusions and Recommendations

In the present study, marker-assisted selection was performed in BC₂F₁ and BC₂F₃ populations, and QTLs (STG 3A and STG 3B) associated with stay-green (STG) characteristics from the donor parent B35 were successfully introgressed into two farmers' preferred

recurrent varieties, NACO Mtama 1 and Seguifa. Nonetheless, populations generated from Pato, Macia, Wagita, Kenya, Tegemeo, Hakika and Wahi backgrounds did not continue due to the poor performance of either STG expression or associated traits such as leaf rolling, grain yield and its related traits. Besides better drought tolerance, improved lines also exhibited higher grain weight. The improved lines generated in the present study will further diversify the base of farmers' preferred sorghum varieties in Tanzania that may help to cope with scarcity of rainfall, especially in semi-arid areas of Sub-Saharan Africa. The introgression lines of sorghum obtained in our study may be used as the bases for climate resilience seed to cope with climate change in Sub-Saharan Africa. Finally, our results suggest that the marker-assisted backcrossing together with field evaluations is sufficient to introgress the target traits from the donor source and recover desirable lines from recurrent parents.

Supplementary Materials: The following are available online at <https://www.mdpi.com/article/10.3390/agriculture11090883/s1>. Table S1: BC₂F₁ sorghum populations scored with heterozygous alleles using SNP markers, Table S2: Genotypes of BC₂F₃ populations scored by favourable alleles using SNP markers.

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References

1. Barron, J.; Rockström, J.; Gichuki, F.; Hatibu, N. Dry spell analysis and maize yields for two semi-arid locations in East Africa. *Agric. Meteorol.* **2003**, *117*, 23–37. [[CrossRef](#)]
2. Chander, S.; Garcia-Oliveira, A.L.; Gedil, M.; Shah, T.; Otusanya, G.O.; Asiedu, R.; Chigeza, G. Genetic diversity and population structure of soybean lines adapted to sub-Saharan Africa using single nucleotide polymorphism (SNP) markers. *Agronomy* **2021**, *11*, 604. [[CrossRef](#)]
3. Premachandra, G.S.; Hahn, D.T.; Joly, R.J. Leaf water relations and gas exchange in two grain sorghum genotypes differing in their pre- and post-flowering drought tolerance. *J. Plant Physiol.* **1994**, *143*, 96–101. [[CrossRef](#)]
4. Kamal, S.M.; Gorafi, Y.S.A.; Tsujimoto, H.; Ghanim, A.M.A. Stay-Green QTLs response in adaptation to post flowering drought depends on the drought severity. *BioMed Res. Int.* **2018**, *2018*, 1–15. [[CrossRef](#)]
5. Borrell, A.K.; van Oosterom, E.J.; Mullet, J.E.; George-Jaeggli, B.; Jordan, D.R.; Klein, P.E.; Hammer, G.L. Stay-green alleles individually enhance grain yield in sorghum under drought by modifying canopy development and water uptake patterns. *New Phytol.* **2014**, *203*, 817–830. [[CrossRef](#)]
6. Rosenow, D.T.; Clark, L.E. Drought tolerance in sorghum. In *Proceedings of the 36th Annual Corn and Sorghum Industry Research Conference*; Loden, H.D., Wilkinson, D., Eds.; American Seed Trade Association: Washington, DC, USA, 1981; pp. 18–31.
7. Slegers, M.F.W. Exploring Farmers' Perceptions of Drought in Tanzania and Ethiopia. Ph.D Thesis, Wageningen University, Wageningen, The Netherlands, 2008.

8. Velazco, J.G.; Jordan, D.R.; Mace, E.S.; Hunt, C.H.; Malosetti, M.; van Eeuwijk, F.A. Genomic Prediction of Grain Yield and Drought-Adaptation Capacity in Sorghum is Enhanced by Multi-Trait Analysis. *Front. Plant Sci.* **2003**, *10*, 997. [[CrossRef](#)]
9. Sory, S. Marker Assisted Selection for Post-Flowering Drought Tolerance in *Sorghum bicolor* [L.] Moench. Ph.D. Thesis, University of Ghana, Accra, Ghana, December 2015.
10. Serraj, R.; Hash, T.C.; Buhariwalla, H.K.; Bidinger, F.R.; Folkertsma, R.T.; Chandra, S.; Gaur, P.; Kashiwagi, J.; Nigam, S.N.; Rupakula, A.; et al. Marker-assisted breeding for crop drought tolerance at ICRISAT: Achievements and prospects. In *Proceedings of the International Congress; In the Wake of the Double Helix: From the Green Revolution to the Gene Revolution*; Tuberosa, R., Phillips, R.L., Gale, M., Eds.; Avenue Media: Bologna, Italy, 2003; pp. 217–238.
11. Boyles, R.E.; Brenton, Z.W.; Kresovich, S. Genetic and genomic resources of sorghum to connect genotype with phenotype in contrasting environments. *Plant J.* **2019**, *97*, 19–39. [[CrossRef](#)]
12. Kamal, N.M.; Gorafi, Y.S.A.; Abdeltwab, H.; Abdalla, I.; Tsujimoto, H.; Ghanim, A.M.A. A New Breeding Strategy towards Introgression and Characterization of Stay-Green QTL for Drought Tolerance in Sorghum. *Agriculture* **2021**, *11*, 598. [[CrossRef](#)]
13. Vadez, V.; Deshpande, S.P.; Kholova, J.; Hammer, G.L.; Borrell, A.K.; Talwar, H.S.; Hash, C.T. Stay-green quantitative trait loci's effects on water extraction, transpiration efficiency and seed yield depend on recipient parent background. *Funct. Plant Biol.* **2011**, *38*, 553–566. [[CrossRef](#)] [[PubMed](#)]
14. Ribaut, J.M.; Betran, J. Single large-scale marker assisted selection (SLS-MAS). *Mol. Breed.* **1999**, *5*, 531–541. [[CrossRef](#)]
15. Xu, W.; Subudhi, P.K.; Crasta, O.R.; Rosenow, D.T.; Mullet, J.E.; Nguyen, H.T. Molecular mapping of QTLs conferring staygreen in grain sorghum (*Sorghum bicolor* L. Moench). *Genome* **2000**, *43*, 461–469. [[CrossRef](#)]
16. Kebede, H.; Subudhi, P.K.; Rosenow, D.T.; Nguyen, H.T. Quantitative trait loci influencing drought tolerance in grain sorghum (*Sorghum bicolor* L. Moench). *Theor. Appl. Genet.* **2001**, *103*, 266–276. [[CrossRef](#)]
17. Ayalew, K.; Adugna, A.; Fetene, M.; Sintayehu, S. Evaluation of bio-physiological and yield responses of stay-green QTL introgression sorghum lines to post-flowering drought stress. *Afr. Crop Sci. J.* **2018**, *26*, 447–462. [[CrossRef](#)]
18. Abdelrahman, M.; El-Sayed, M.; Jogaiah, S.; Burritt, D.J.; Tran, L.P. The “STAY-GREEN” trait and phytohormone signaling networks in plants under heat stress. *Plant Cell Rep.* **2017**, *36*, 1009–1025. [[CrossRef](#)] [[PubMed](#)]
19. Ngugi, K.; Kimani, W.; Kiambi, D.; Mutitu, E.W. Improving Drought Tolerance in *Sorghum bicolor* L. Moench: Marker-Assisted transfer of the Stay-Green Quantitative Trait Loci (QTL) from a Characterized Donor Source into a Local Farmer Variety. *Int. J. Sci.* **2013**, *1*, 154–162. [[CrossRef](#)]
20. Gimode, D.; Odeny, D.A.; de Villiers, E.P.; Wanyonyi, S.; Dida, M.M.; Mneney, E.E.; Muchugi, A.; Machuka, J.; de Villiers, S.M. Identification of SNP and SSR Markers in Finger Millet Using Next Generation Sequencing Technologies. *PLoS ONE* **2016**, *11*, e0159437. [[CrossRef](#)]
21. Mammadov, J.; Aggarwal, R.; Buyyarapu, R.; Kumpatla, S. SNP markers and their impact on plant breeding. *Int. J. Plant Genom.* **2012**, *2012*, 728398. [[CrossRef](#)] [[PubMed](#)]
22. Semagn, K.; Babu, R.; Hearne, S.; Olsen, M. Single nucleotide polymorphism genotyping using Kompetitive Allele Specific PCR (KASP): Overview of the technology and its application in crop improvement. *Mol. Breed.* **2013**, *33*, 1–14. [[CrossRef](#)]
23. Thomson, M.J. High-throughput SNP genotyping to accelerate crop improvement. *Plant Breed Biotech.* **2014**, *2*, 195–212. [[CrossRef](#)]
24. Meade, F.; Byrne, S.; Griffin, D.; Kennedy, C.; Mesiti, F.; Dan Millbourne, D. Rapid Development of KASP Markers for Disease Resistance Genes Using Pooled Whole-Genome Re-sequencing. *Potato Res.* **2020**, *63*, 57–73. [[CrossRef](#)]
25. Kiranmayee, K.N.S.U.; Hash, C.T.; Sivasubramani, S.; Ramu, P.; Amindala, B.P.; Rathore, A.; Kishor, P.B.K.; Gupta, R.; Deshpande, S.P. Fine-Mapping of Sorghum Stay-Green QTL on Chromosome10 Revealed Genes Associated with Delayed Senescence. *Genes* **2020**, *11*, 1026. [[CrossRef](#)] [[PubMed](#)]
26. Deshpande, S.; Rakshit, S.; Manasa, K.G.; Pandey, S.; Gupta, R. Genomic Approaches for Abiotic Stress Tolerance in Sorghum. In *The Sorghum Genome*; Springer: Cham, Switzerland, 2016; pp. 169–187.
27. Meru, G.M. Genotyping BC₃F₂ Populations of Four Ethiopian Sorghum Varieties for Stay Green QTLs Introgression through Marker Assisted Selection with SSRs. Master's Thesis, Kenyatta University, Nairobi, Kenya, 2010.
28. International Board for Plant Genetic Resources (IBPGR); International Crops Research Institute for the Semi-Arid Tropics (ICRISAT). *Descriptors for Sorghum [Sorghum bicolor (L.) Moench]*; IBPGR: Rome, Italy; ICRISAT: Patancheru, India, 1993.
29. Burow, G.; Chopra, R.; Sattler, S.; Burke, J.; Acosta-Martinez, V.; Xin, Z. Deployment of SNP (CAPS and KASP) markers for allelic discrimination and easy access to functional variants for brown midrib genes bmr6 and bmr12 in *Sorghum bicolor*. *Mol. Breed.* **2019**, *39*, 115. [[CrossRef](#)]
30. Da Silva, M.J.; Ribeiro, P.C.O.; Silva, R.A.; Silva, K.J.; Schaffert, R.E.; Parrella, R.A.C. Multi-trait selection of sweet sorghum (*Sorghum bicolor* (L.) Moench) genotypes for bioenergy production. *J. Bioenergy Food Sci.* **2020**, *7*, e2952020. [[CrossRef](#)]
31. Schertz, K.F.; Clark, L.E. Controlling dehiscence with plastic bags for hand crosses in Sorghum 1. *Crop Sci.* **1967**, *7*, 540–542. [[CrossRef](#)]
32. Sakuraba, Y.; Park, S.; Kim, Y.; Wang, S.; Yoo, S.; Hörtensteiner, S.; Paek, N. *Arabidopsis* STAY-GREEN2 Is a Negative Regulator of Chlorophyll Degradation during Leaf Senescence. *Mol. Plant* **2014**, *7*, 1288–1302. [[CrossRef](#)]
33. Sukumaran, S.; Li, X.; Xianran Li, X.; Zhu, C.; Bai, G.; Perumal, R.; Tuinstra, M.R.; Prasad, P.V.; Mitchell, S.E.; Tesso, T.T.; et al. QTL Mapping for Grain Yield, Flowering Time, and Stay-Green Traits in Sorghum with Genotyping-by-Sequencing Markers. *Crop Sci.* **2016**, *56*, 1429–1442. [[CrossRef](#)]

34. Kimani, W.; Ngugi, K.; Kiambi, D.; Mutitu, E.W.; De Villiers, S. Marker-assisted introgression of stay-green trait in a Kenyan sorghum variety. *E. Afr. Agric. For. J.* **2012**, *78*, 109–115.
35. Reddy, N.R.; Ragimasalawada, M.; Sabbavarapu, M.M.; Nadoor, S.; Patil, J.V. Detection and validation of stay-green QTL in post-rainy sorghum involving widely adapted cultivar, M35-1 and a popular stay-green genotype B35. *BMC Genom.* **2014**, *15*, 909. [[CrossRef](#)]
36. Kamal, N.M.; Gorafi, Y.S.A.; Ghanim, A.M.A. Performance of Sorghum stay-green introgression lines under post-flowering drought. *Int. J. Plant Res.* **2017**, *7*, 65–74.
37. Kholova, J.; Deshpande, S.; Madhusudhana, R.; Blummel, M.; Borrell, A.; Hammer, G. *Improving Post-Rainy Sorghum Varieties to Meet the Growing Grain and Fodder Demand in India—Phase 2*; Australian Centre for International Agricultural Research: Canberra, Australia, 2019; p. 29.
38. Adu-Boakyewaa, G.A.; Badu-Apraku, B.; Akromah, R.; Garcia-Oliveira, A.L.; Awuku, F.J.; Gedil, M. Genetic diversity and population structure of early-maturing tropical maize inbred lines using SNP markers. *PLoS ONE* **2019**, *14*, e021481.
39. Nelimor, C.; Badu-Apraku, B.; Garcia-Oliveira, A.L.; Tetteh, A.; Paterne, A.; N'guetta, A.S.; Gedil, M. Genomic Analysis of Selected Maize Landraces from Sahel and Coastal West Africa Reveals Their Variability and Potential for Genetic Enhancement. *Genes* **2020**, *11*, 1054. [[CrossRef](#)] [[PubMed](#)]
40. Platten, J.D.; Cobb, J.N.; Zantua, R.E. Criteria for evaluating molecular markers: Comprehensive quality metrics to improve marker-assisted selection. *PLoS ONE* **2019**, *14*, e0210529. [[CrossRef](#)]
41. Azzouz-Olden, F.; Hunt, A.G.; Dinkins, R. Transcriptome analysis of drought-tolerant sorghum genotype SC56 in response to water stress reveals an oxidative stress defense strategy. *Mol. Biol. Rep.* **2020**, *47*, 3291–3303. [[CrossRef](#)]